Functional roles of exosomes in cardiovascular disorders: a systematic review

H.-J. SUN, X.-X. ZHU, W.-W. CAI, L.-Y. QIU

Department of Basic Medicine, Wuxi Medical School, Jiangnan University, Wuxi, Jiangsu, P.R. China

Abstract. - Cardiovascular diseases are major causes of people death associated with high mortality and disability. Exosomes are nanosized extracellular vesicles containing protein, lipid, transcription factors, mRNAs, non-coding RNA (ncRNA) and nucleic acid contents, which are critical players of intercellular communication via long-range signals or cell-to-cell contact. The emergence of exosomes provides favorable strategies for the diagnosis and treatment of cardiovascular diseases. Exosomes-based molecular mechanisms are important for developing novel therapeutic approaches for cardiovascular events. In this review, we will (1) provide insights into the detrimental and beneficial effects of exosomes on cardiovascular physiology, (2) summarize the underlying biological mechanisms of the exosome in cardiovascular events, (3) investigate the therapeutic value of exosomes for cardiovascular disorders.

Key Words:

Exosomes, Cardiovascular diseases, Endothelial cell.

Introduction

The prevalence of cardiovascular diseases is markedly increased in low- and middle-income countries for decades^{1,2}. Over 4.3 million deaths are induced by cardiovascular diseases every year in Europe, which brings a considerable burden on the economy of European Union³. The Centers for Disease Control and Prevention have announced that \$444 billion may be used for the treatment of cardiovascular diseases in 2010, and the costs will be enhanced with the increase of life expectancy⁴. Development of novel diagnostic or therapeutic strategies may provide multiple opportunities for reduction in mortality of cardiovascular diseases.

Exosomes have obtained substantially attention due to their potential therapeutic applications⁵. A wide range of researches has investigated the

roles of exosomes in cancers^{6,7}, neurologic disorder⁸, endocrine system diseases⁹, autoimmune diseases¹⁰ and cardiovascular diseases¹¹. Exosomes are involved in various biological activities including cell proliferation and differentiation¹², inflammation¹³, senescence¹⁴, angiogenesis¹⁵, stress response¹⁶ and cardiovascular remodeling¹⁷. Exosomes-mediated intercellular communication plays a fundamental role in vascular integrity and cardiovascular diseases¹⁸.

Exosomes are associated with many cardiovascular pathologies such as cardiac hypertrophy¹⁹, atherogenesis²⁰, heart failure²¹, hypertension²² and diabetic cardiomyopathy²³. Mounting evidence has shown that exosomes may transfer non-coding RNA (ncRNA) including miRNA and lncRNA to recipient cells, thus leading to the changes in protein expressions and phenotypes of recipient cells^{24,25}. Exosomes are recently used as disease biomarkers²⁶, therapeutic targets²⁷, agents for drug delivery²⁸ and biomedical applications²⁹. The following review will summarize the intercellular signaling, possible mechanisms, prognostic, diagnostic and therapeutic roles of exosomes and exosomal ncRNAs in cardiovascular diseases.

Biogenesis and Secretion of Exosome

Cell to cell communication between cardio-vascular cells is a complex process that exerts a requisite role in cardiovascular biology^{30,31}. Accumulating evidence establishes that exosomes are intercellular communication messengers^{32,33}. The exosomes were firstly identified during the research on the formation of vesicle in 1987³⁴. Exosomes are known to be one of the subtypes of membrane vesicles, whose sizes are ranging from 30 to 100 nm³⁵. Exosomes are distinguished from apoptotic bodies and microvesicles due to their unique qualities³⁶.

It has been demonstrated that microvesicles are released from direct outward blebs of plasma. However, exosomes are produced by en-

dosomal network³⁷. The inward budding of cell membrane ligands leads to the fusion of small vesicles and early endosomes. The extracellular membrane ligands are internalized to surfaces of these small vesicles during this process. The second inward invagination of the endocytic vesicles membrane creates various intraluminal vesicles (late endosomes). The deposition of late endosomes is defined as multivesicular bodies. The multivesicular bodies are then fused into the cell membrane, following by release of intraluminal vesicles through an exocytotic way. The released intraluminal vesicles are referred to as exosomes. A wide coverage of cargos such as proteins, enzymes, ncRNA, mRNA, and molecules are presented within exosomes^{23,38}.

The constitutive or inducible pathways are responsible for the release of exosomes. In the literature, certain RAB GTPases³⁹⁻⁴¹, WNT5A⁴², heterotrimeric G-protein⁴³, glycosphingolipids and flotillins⁴⁴ can modulate the constitutive secretion of exosomes. Numerous factors including calcium release-dependent mechanism⁴⁵, heat shock⁴⁶, hypoxia⁴⁷, thrombin⁴⁸, DNA damage⁴⁹, lipopolysaccharide^{50,51} participate in the secretion of exosomes.

Characterization of Exosomes

Electron microscopy is a critical step in the characterization of exosomes. Transmission electron microscopy can clearly capture the photographs of exosomes with the aid of uranyl acetate and methylcellulose. Exosomes are observed as double-membrane bound vesicles under electron microscopy⁵². The "cup-shaped" morphology of exosomes can be distinguished on electron micrograph⁵³. Furthermore, standard preparation techniques are applied to identify exosomes on tissues using electron micrographs⁵⁴.

It is noted that exosomes are generated from endosomal pathways, antibodies against endosomal markers may be employed to characterize the exosomes. Tetraspanins (CD9, CD63, and CD81), and phosphatidylserine are abundantly expressed within exosomes⁵⁵. Combinations of antibodies and electron micrograph methods are recommended to obtain accurate confirmation of exosomes

Flow cytometry is applied to examine fluorophores-tagged exosomes, but it is unable to quantify the exosome numbers due to swarming effects^{56,57}. The exosomes are marked by membrane-binding dye such as PKH67, which can be seen under fluorescence and confocal micros-

copy. Such technics could determine whether marked exosomes are absorbed into recipient cells⁵⁸. Moreover, small-angle X-ray scattering⁵⁹, resistive pulse sensing⁶⁰, and Raman microspectroscopy⁶¹ are novel methods for detection of exosomes.

Cellular Communication Functions

Cell junctions, adhesion contacts, and soluble factors are classical molecules, and they act on targeted cells in an endocrine manner⁶². Extracellular vesicles transfer the various proteins, lipids, and nucleic acids into recipient cells, thus causing changes in intracellular signaling of recipient cells⁵². A growing body of evidence indicates that the proteins, mRNA, miRNA and lnc RNA within exosomes are inserted into recipient cells, thus inducing transient or persistent phenotypic changes in recipient cells⁶³. It is interesting that the small RNAs in the exosomes are surrounded by lipids or lipoprotein complexes, which may protect them from degradation during the transport processes⁶⁴. The exosomes are involved in various physiological or pathological processes such as regulation of tumor growth, cytokine production or cardiovascular disorders^{9,65,66}.

Biomarkers, Diagnosis, and Therapy of Exosomes

With the deepening of research on exosomes, the exosomes may be served as valuable biomarkers, diagnostic, prognostic and therapeutic tools for cardiovascular diseases^{67,68}. MiR-133a-containing exosomes are a useful biomarker for myocardial damage or cardiomyocyte death⁶⁹. It is revealed that the levels of miR-15b, miR-34a, and miR-636 within urinary exosomes are enhanced in patients with type 2 diabetic kidney disease, and these urinary exosomal miRs are treated as a novel diagnostic panel for diabetic kidney disease⁷⁰. Bioinformatics analysis establishes that urinary exosomal miR-133b, miR-342 and miR-30a are closely associated with systolic-diastolic blood pressure, serum creatinine, urinary albumin creatinine ratio and glomerular filtration rate in diabetic nephropathy⁷¹.

The biomolecules and bioactive molecules such as proteins, enzymes, growth factors, mR-NA, DNA, and ncRNAs in exosomes facilitate the exosomes to be a therapeutic tool in many diseases⁷². In addition, exosomes are chemically modified to be a delivery tool for transferring the specific bioactive molecules into certain cell types⁷³. The exosomes-carrying tumor antigens

induce T-cell lymphocyte responses and inhibit tumor growth⁷⁴. The potential roles of exosomes in cardiovascular diseases are intensively investigated in recent years. The exosomes derived from dendritic cells stimulate CD4(+) T lymphocytes activation to improve cardiac function after myocardial infarction in mice⁷⁵. The cardiomyocyte-released exosomes transfer glucose transport to endothelial cells, thus inducing glucose uptake, glycolytic activity, and pyruvate production in endothelium⁷⁶. Mesenchymal stem cells (MSCs) overexpressing GATA-4 releases exosomes containing a reservoir of anti-apoptotic microRNAs to rat neonatal cardiomyocytes, contributing to cardiomyocytes survival under hypoxic environment⁷⁷.

To date, the possible roles of exosomes in cardiovascular diseases have not yet been fully elucidated in the clinical practice. More and more studies should be conducted to examine diagnostic, prognostic value and functional roles of exosomes content in cardiovascular diseases.

Exosomes and Diabetes Mellitus

Diabetes mellitus is a widely prevalent disorder around the world78,79. The exosomes are closely associated with diabetes in diabetic patients or diabetes models80-88. Plasma exosomal miR-326 levels are up-regulated, but let-7a and let-7f levels are down-regulated in diabetic patients, the levels of let-7a and let-7f in plasma exosomes are significantly increased after antidiabetic treatment⁸¹. The cardiomyocyte-derived exosomes from diabetic rats inhibit the proliferation and migration of endothelial cells, but the exosomes from normal rats accelerate the proliferation and migration of endothelial cells⁸⁸. It has been recently reported that the cardiomyocytes-derived exosomes contribute to increases in glucose uptake, glycolysis in endothelial cells under glucose deprivation conditions⁷⁶. The cardiomyocytes transfer the exosomal miR-320 into endothelial cells to mediate angiogenesis in type 2 diabetic rats⁸³. The exosomes from bone marrow-derived mesenchymal stem cells are transferred into damaged neurons and astrocytes, which significantly improved cognitive impairment in diabetic mice⁸⁹. A large prospective study has concluded that exosomes containing miR-126 have a predictive value for cardiovascular events in patients with stable coronary artery disease⁹⁰. The endothelial cells-derived exosomes promote vascular endothelial repair via transferring the miR-126 into recipient cells, which

is disrupted under hyperglycemic conditions⁹¹. The miRNA-enriched exosomes from fibrocytes accelerate wound healing in diabetic mice⁹². The exosomes are ideal candidates for illumination of diabetic pathophysiology, and may provide novel therapeutic approaches for diabetes.

Exosomes and Myocardial Infarction

Myocardial infarction is reflected by occlusion of coronary vessels and cardiac cell death^{93,94}. The molecule mechanisms for cardiac rehabilitate response to myocardial infarction are not fully explained95. Coronary bypass surgery and balloon dilatation of coronary vessels are usually used to alleviate cardiac impairment in the acute phase of myocardial infarction⁹⁶. Novel strategies or technics are urgent to be developed for improvement of cardiac tissue repair. The exosomes are critically involved in the proliferation and apoptosis of targeted cells⁹⁷. A plethora of researches has identified the roles of exosomes in cardiovascular diseases98-100. The exosomes are essential for local and distant microcommunication with recipient cells in myocardial infarction^{12,101}. The cardiac progenitor cells¹⁰² or embryonic stem cells-releases exosomes¹⁰³ regulate cardiac regeneration and cardiac remodeling during the myocardial infarction.

Mesenchymal stem cells are able to deliver miR-22-shutting exosomes into neonatal rat ventricle cardiomyocytes, leading to reduced apoptosis of cardiomyocytes¹⁰⁴. Cardiac progenitor cells contribute to decreased cardiac fibrosis, cardiomyocyte apoptosis, and increased angiogenesis or cardiac output after myocardial infarction via transferring antifibrotic miRNAs-enriched exosomes to fibroblasts under hypoxia^{32,102}. The cardiosphere-released exosomes stimulate the proliferation and angiogenesis of cardiomyocytes¹⁰⁵. The mesenchymal stem cell-derived exosomes preserve cardiac function, and relieve infarct size in ischemia reperfusion injury mode¹⁰⁶. Intravenous administration of mesenchymal stem cells-derived exosomes decreases the infarct size by 45% and depresses systemic inflammation in ischemia-reperfusion model¹⁰⁷. The exosomes from healthy controls exert a protective role in ischemic myocardium via delivering endogenous protective signals including cardio-protective heat shock protein 70¹⁰⁸. Direct intramyocardial transplantation of exosomes from GATA-4 overexpressed mesenchymal stem cells obviously improve cardiac contractile function and alleviate infarct size in the rat heart⁷⁷. These studies suggest that exosomes from stem cells are believed to play protective roles in cardiac remodeling during the myocardial infarction.

Exosomes and Coronary Artery Disease

Atherosclerotic lesions are closely associated with endothelial cell activation, inflammation, formation of foam cells and phenotype transformation of VSMCs^{109,110}. In primary rat aortic endothelial cells, the heat shock protein-70-carrying exosomes are increased in response to homocysteine and ox-LDL stimulation¹¹¹. Heat shock protein-70 mediated proinflammatory genes contribute to monocyte adhesion in endothelial cells¹¹². The heat shock protein-70-enriching exosomes may be responsible for subendothelial migration of monocytes in atherosclerosis. The activated macrophages secrete miR-223-containg exosomes to evoke an inflammatory response in atherosclerosis¹¹³. It has been shown that exosomes from atherosclerotic plagues are a stimulator for the adhesion molecule expressions, and inflammatory endothelial cells, which may be responsible for the plaque development¹¹⁴. The exosomes containing miR-143/145 are increased in human umbilical vein endothelial cells exposure to shear stress through modulation of shear-responsive transcription factor KLF2¹¹⁵⁻¹¹⁷. Cardiomyocytes and endothelial cells can communicate via exosomes-mediated exchanges^{118,119}. Endothelial cells release miR-146a-bearing exosomes to cardiomyocytes, which downregulates the interleukin-1 receptor-associated kinase 1 and receptor tyrosine-protein kinase ERBB4 levels in cardiomyocytes 118,120.

Activated platelets-derived exosomes carry CD40 ligand to regulate the differentiation of antigen-presenting cells including monocyte-derived dendritic cells¹²¹. However, stored platelets-associated exosomes retard the differentiation from monocytes to macrophage and dendritic cell maturation¹²². It is seen that platelet-released exosomes may exert different effects on inflammation response. Also, the platelet-derived exosomes may participate in atherogenesis via hyperplasia of vascular smooth muscle cells¹²³ and proinflammatory activation of endothelial cells¹²⁴. The monocytes-generated exosomes promote atherogenesis associated with activation of macrophages and endothelial cells¹²⁵. The monocytes-derived exosomes are suggested to stimulate nitrosative stress in human endothelial cells¹²⁶.

Conclusions

In recent years, the exosomes are novel approaches or strategies for characterizing the communications between living cells. The functional roles of exosomes in cardiovascular disorders are summarized in Figure 1. The exosomes are taken as possible candidates for intercellular and tissue-level communication. Importantly, the exosomes-containing various proteins and RNA messages may be secreted to recipient cells, which modulates the targeted gene expressions in recipient cells. Furthermore, the epigenetic mechanisms such as histone modifications, DNA methylation, and non-coding RNA expressions play pivotal roles in various biological effects in cardiovascular diseases. It may be speculated that exosomes may carry epigenetic modulator to induce functional changes in recipient cells. It is interesting that exosomes from different cells may exhibit protective or destructive roles in cardiovascular diseases. The advanced technics to modify or load thera-

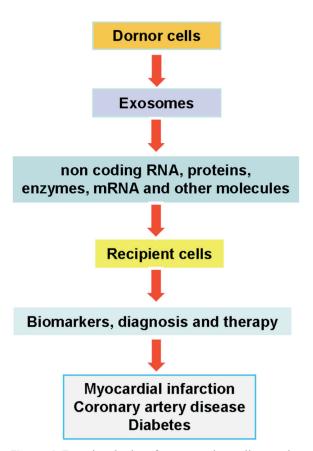


Figure 1. Functional roles of exosomes in cardiovascular disorders.

peutics into exosomes can be developed and standardized in a future study. It is undeniable that the unique opportunities and new challenges for characterization of exosomes as clinical biomarkers, diagnosis and prognosis factors in cardiovascular diseases are still on fire.

Acknowledgements

This work was supported in part by grants from Fundamental Research Funds for the Central Universities (grant no. JUSRP51412B).

Conflict of Interest

The Authors declare that they have no conflict of interests.

References

- OFORI SN, ODIA OJ. Risk assessment in the prevention of cardiovascular disease in low-resource settings. Indian Heart J 2016; 68: 391-398.
- Fu DG. Regulation of redox signalling and autophagy during cardiovascular diseases-role of resveratrol. Eur Rev Med Pharmacol Sci 2015; 19: 1530-1536.
- 3) FUSTER V, KELLY BB, VEDANTHAN R. Promoting global cardiovascular health: moving forward. Circulation 2011; 123: 1671-1678.
- 4) HENDRANI AD, ADESIYUN T, QUISPE R, JONES SR, STONE NJ, BLUMENTHAL RS, MARTIN SS. Dyslipidemia management in primary prevention of cardiovascular disease: Current guidelines and strategies. World J Cardiol 2016; 8: 201-210.
- XITONG D, XIAORONG Z. Targeted therapeutic delivery using engineered exosomes and its applications in cardiovascular diseases. Gene 2016; 575: 377-384.
- ZHANG X, PEI Z, CHEN J, JI C, XU J, ZHANG X, WANG J. Exosomes for Immunoregulation and Therapeutic Intervention in Cancer. J Cancer 2016; 7: 1081-1087.
- Xu CG, Yang MF, Ren YQ, Wu CH, Wang LQ. Exosomes mediated transfer of IncRNA UCA1 results in increased tamoxifen resistance in breast cancer cells. Eur Rev Med Pharmacol Sci 2016; 20: 4362-4368.
- TSILIONI I, PANAGIOTIDOU S, THEOHARIDES TC. Exosomes in neurologic and psychiatric disorders. Clin Ther 2014; 36: 882-888.
- LAWSON C, VICENCIO JM, YELLON DM, DAVIDSON SM. Microvesicles and exosomes: new players in metabolic and cardiovascular disease. J Endocrinol 2016; 228: R57-71.
- TURPIN D, TRUCHETET ME, FAUSTIN B, AUGUSTO JF, CONTIN-BORDES C, BRISSON A, BLANCO P, ET AL. Role of extracellular vesicles in autoimmune diseases. Autoimmun Rev 2016; 15: 174-183.

- PFEIFER P, WERNER N, JANSEN F. Role and function of MicroRNAs in extracellular vesicles in cardiovascular biology. Biomed Res Int 2015; 2015: 161393.
- 12) SINGLA DK. Stem cells and exosomes in cardiac repair. Curr Opin Pharmacol 2016; 27: 19-23.
- Bonjoch L, Casas V, Carrascal M, Closa D. Involvement of exosomes in lung inflammation associated with experimental acute pancreatitis. J Pathol 2016; 240: 235-245.
- 14) VAN BALKOM BW, DE JONG OG, SMITS M, BRUMMELMAN J, DEN OUDEN K, DE BREE PM, VAN EUNDHOVEN MA, PEGTEL DM, STOORVOGEL W, WÜRDINGER T, VERHAAR MC. Endothelial cells require miR-214 to secrete exosomes that suppress senescence and induce angiogenesis in human and mouse endothelial cells. Blood 2013; 121: 3997-4006, s3991-3915.
- DENG Z, RONG Y, TENG Y, ZHUANG X, SAMYKUTTY A, MU J, ZHANG L, CAO P, YAN J, MILLER D, ZHANG HG. Exosomes miR-126a released from MDSC induced by DOX treatment promotes lung metastasis. Oncogene 2017; 36: 639-651.
- 16) Shu M, Taddeo B, Roizman B. Tristetraprolin recruits the herpes simplex virion host shutoff RNase to AU-rich elements in stress response mRNAs to enable their cleavage. J Virol 2015; 89: 5643-5650.
- 17) LYU L, WANG H, LI B, QIN Q, QI L, NAGARKATTI M, NAGARKATTI P, JANICKI JS, WANG XL, CUI T. A critical role of cardiac fibroblast-derived exosomes in activating renin angiotensin system in cardiomyocytes. J Mol Cell Cardiol 2015; 89: 268-279.
- 18) MIRZAPOIAZOVA T, LENNON FE, MAMBETSARIEV B, ALLEN M, RIEHM J, POROYKO VA, SINGLETON PA. Extracellular vesicles from caveolin-enriched microdomains regulate hyaluronan-mediated sustained vascular integrity. Int J Cell Biol 2015; 2015: 481493.
- 19) BANG C, BATKAI S, DANGWAL S, GUPTA SK, FOINQUINOS A, HOLZMANN A, JUST A, REMKE J, ZIMMER K, ZEUG A, PONIMASKIN E, SCHMIEDL A, YIN X, MAYR M, HALDER R, FISCHER A, ENGELHARDT S, WEI Y, SCHOBER A, FIEDLER J, THUM T. Cardiac fibroblast-derived microRNA passenger strand-enriched exosomes mediate cardiomyocyte hypertrophy. J Clin Invest 2014; 124: 2136-2146.
- 20) GIOIA M, VINDIGNI G, TESTA B, RANIOLO S, FASCIGLIONE GF, COLETTA M, BIOCCA S. Membrane cholesterol modulates LOX-1 shedding in endothelial cells. PLoS One 2015; 10: e0141270.
- CAMPBELL CR, BERMAN AE, WEINTRAUB NL, TANG YL. Electrical stimulation to optimize cardioprotective exosomes from cardiac stem cells. Med Hypotheses 2016; 88: 6-9.
- 22) VAN BALKOM BW, EISELE AS, PEGTEL DM, BERVOETS S, VERHAAR MC. Quantitative and qualitative analysis of small RNAs in human endothelial cells and exosomes provides insights into localized RNA processing, degradation and sorting. J Extracell Vesicles 2015; 4: 26760.
- AILAWADI S, WANG X, GU H, FAN GC. Pathologic function and therapeutic potential of exosomes in cardiovascular disease. Biochim Biophys Acta 2015; 1852: 1-11.

- 24) JOYCE DP, KERIN MJ, DWYER RM. Exosome-encapsulated microRNAs as circulating biomarkers for breast cancer. Int J Cancer 2016; 139: 1443-1448.
- LUDWIG AK, GIEBEL B. Exosomes: small vesicles participating in intercellular communication. Int J Biochem Cell Biol 2012; 44: 11-15.
- 26) HIGGINBOTHAM JN, ZHANG Q, JEPPESEN DK, SCOTT AM, MANNING HC, OCHIENG J, FRANKLIN JL, COFFEY RJ. Identification and characterization of EGF receptor in individual exosomes by fluorescence-activated vesicle sorting. J Extracell Vesicles 2016; 5: 29254.
- MATSUSHITA H, YANG YM, PANDOL SJ, SEKI E. Exosome migration inhibitory factor as a marker and therapeutic target for pancreatic cancer. Gastroenterology 2016; 150: 1033-1035.
- 28) LI L, ZHANG L, KNEZ M. Comparison of two endogenous delivery agents in cancer therapy: exosomes and ferritin. Pharmacol Res 2016; 110: 1-9.
- 29) SALIDO-GUADARRAMA I, ROMERO-CORDOBA S, PERAL-TA-ZARAGOZA O, HIDALGO-MIRANDA A, RODRIGUEZ-DOR-ANTES M. MicroRNAs transported by exosomes in body fluids as mediators of intercellular communication in cancer. Onco Targets Ther 2014; 7: 1327-1338.
- 30) KISHORE R, KHAN M. More than tiny sacks: stem cell exosomes as cell-free modality for cardiac repair. Circ Res 2016; 118: 330-343.
- 31) RAFII S, BUTLER JM, DING BS. Angiocrine functions of organ-specific endothelial cells. Nature 2016; 529: 316-325.
- 32) BARILE L, LIONETTI V, CERVIO E, MATTEUCCI M, GHER-GHICEANU M, POPESCU LM, TORRE T, SICLARI F, MOCCETTI T, VASSALLI G. Extracellular vesicles from human cardiac progenitor cells inhibit cardiomyocyte apoptosis and improve cardiac function after myocardial infarction. Cardiovasc Res 2014; 103: 530-541.
- 33) VAN DONGEN HM, MASOUMI N, WITWER KW, PEGTEL DM. Extracellular vesicles exploit viral entry routes for cargo delivery. Microbiol Mol Biol Rev 2016; 80: 369-386.
- 34) JOHNSTONE RM, ADAM M, HAMMOND JR, ORR L, TURBIDE C. Vesicle formation during reticulocyte maturation. Association of plasma membrane activities with released vesicles (exosomes). J Biol Chem 1987; 262: 9412-9420.
- 35) VLASSOV AV, MAGDALENO S, SETTERQUIST R, CONRAD R. Exosomes: current knowledge of their composition, biological functions, and diagnostic and therapeutic potentials. Biochim Biophys Acta 2012; 1820: 940-948.
- 36) RECORD M, CARAYON K, POIROT M, SILVENTE-POIROT S. Exosomes as new vesicular lipid transporters involved in cell-cell communication and various pathophysiologies. Biochim Biophys Acta 2014; 1841: 108-120.
- 37) ALENQUER M, AMORIM MJ. Exosome biogenesis, regulation, and function in viral infection. Viruses 2015; 7: 5066-5083.

- MILLER IV, GRUNEWALD TG. Tumour-derived exosomes: tiny envelopes for big stories. Biol Cell 2015; 107: 287-305.
- 39) OSTROWSKI M, CARMO NB, KRUMEICH S, FANGET I, RAPOSO G, SAVINA A, MOITA CF, SCHAUER K, HUME AN, FREITAS RP, GOUD B, BENAROCH P, HACOHEN N, FUKUDA M, DESNOS C, SEABRA MC, DARCHEN F, AMIGORENA S, MOITA LF, THERY C. Rab27a and Rab27b control different steps of the exosome secretion pathway. Nat Cell Biol 2010; 12: 19-30; sup pp 11-13.
- SAVINA A, VIDAL M, COLOMBO MI. The exosome pathway in K562 cells is regulated by Rab11. J Cell Sci 2002; 115: 2505-2515.
- 41) HSU C, MOROHASHI Y, YOSHIMURA S, MANRIQUE-HOYOS N, JUNG S, LAUTERBACH MA, BAKHTI M, GRØNBORG M, MÖBIUS W, RHEE J, BARR FA, SIMONS M. Regulation of exosome secretion by Rab35 and its GTPase-activating proteins TBC1D10A-C. J Cell Biol 2010; 189: 223-232.
- 42) EKSTROM EJ, BERGENFELZ C, VON BULOW V, SERIFLER F, CARLEMALM E, JONSSON G, ANDERSSON T, LEANDERSSON K. WNT5A induces release of exosomes containing pro-angiogenic and immunosuppressive factors from malignant melanoma cells. Mol Cancer 2014; 13: 88.
- 43) Noguchi E, Hayashi N, Azuma Y, Seki T, Nakamura M, Nakashima N, Yanagida M, He X, Mueller U, Sazer S, Nishimoto T. Dis3, implicated in mitotic control, binds directly to Ran and enhances the GEF activity of RCC1. EMBO J 1996; 15: 5595-5605.
- 44) PHUYAL S, HESSVIK NP, SKOTLAND T, SANDVIG K, LLORENTE A. Regulation of exosome release by glycosphingolipids and flotillins. FEBS J 2014; 281: 2214-2227.
- 45) SAVINA A, FURLAN M, VIDAL M, COLOMBO MI. Exosome release is regulated by a calcium-dependent mechanism in K562 cells. J Biol Chem 2003; 278: 20083-20090.
- 46) CHEN T, GUO J, YANG M, ZHU X, CAO X. Chemokine-containing exosomes are released from heat-stressed tumor cells via lipid raft-dependent pathway and act as efficient tumor vaccine. J Immunol 2011; 186: 2219-2228.
- 47) PARK JE, TAN HS, DATTA A, LAI RC, ZHANG H, MENG W, LIM SK, SZE SK. Hypoxic tumor cell modulates its microenvironment to enhance angiogenic and metastatic potential by secretion of proteins and exosomes. Mol Cell Proteomics 2010; 9: 1085-1099.
- 48) HEUNEN HF, SCHIEL AE, FINHEER R, GEUZE HJ, SIXMA JJ. Activated platelets release two types of membrane vesicles: microvesicles by surface shedding and exosomes derived from exocytosis of multivesicular bodies and alpha-granules. Blood 1999; 94: 3791-3799.
- 49) Lespagnol A, Duflaut D, Beekman C, Blanc L, Fiucci G, Marine JC, Vidal M, Amson R, Telerman A. Exosome secretion, including the DNA damage-induced p53-dependent secretory pathway, is severely compromised in TSAP6/Steap3-null mice. Cell Death Differ 2008; 15: 1723-1733.

- 50) GAMBIM MH, DO CARMO ADE O, MARTI L, VERISSIMO-FIL-HO S, LOPES LR, JANISZEWSKI M. Platelet-derived exosomes induce endothelial cell apoptosis through peroxynitrite generation: experimental evidence for a novel mechanism of septic vascular dysfunction. Crit Care 2007; 11: R107.
- 51) MALIK ZA, KOTT KS, POE AJ, KUO T, CHEN L, FERRARA KW, KNOWLTON AA. Cardiac myocyte exosomes: stability, HSP60, and proteomics. Am J Physiol Heart Circ Physiol 2013; 304: H954-965.
- THERY C, OSTROWSKI M, SEGURA E. Membrane vesicles as conveyors of immune responses. Nat Rev Immunol 2009; 9: 581-593.
- 53) VAN WEERING JR, BROWN E, SHARP TH, MANTELL J, CUL-LEN PJ, VERKADE P. Intracellular membrane traffic at high resolution. Methods Cell Biol 2010; 96: 619-648.
- 54) FEVRIER B, RAPOSO G. Exosomes: endosomal-derived vesicles shipping extracellular messages. Curr Opin Cell Biol 2004; 16: 415-421.
- 55) Denzer K, Kleijmeer MJ, Heijnen HF, Stoorvogel W, Geuze HJ. Exosome: from internal vesicle of the multivesicular body to intercellular signaling device. J Cell Sci 2000; 19: 3365-3374.
- 56) Dragovic RA, Gardiner C, Brooks AS, Tannetta DS, Ferguson DJ, Hole P, Carr B, Redman CW, Harris AL, Dobson PJ, Harrison P, Sargent IL. Sizing and phenotyping of cellular vesicles using Nanoparticle Tracking Analysis. Nanomedicine 2011; 7: 780-788.
- 57) VAN DER POL E, VAN GEMERT MJ, STURK A, NIEUWLAND R, VAN LEEUWEN TG. Single vs. swarm detection of microparticles and exosomes by flow cytometry. J Thromb Haemost 2012; 10: 919-930.
- 58) Lugini L, Cecchetti S, Huber V, Luciani F, Macchia G, Spadaro F, Paris L, Abalsamo L, Colone M, Molinari A, Podo F, Rivoltini L, Ramoni C, Fais S. Immune surveillance properties of human NK cell-derived exosomes. J Immunol 2012; 189: 2833-2842.
- 59) VAN DER POL E, COUMANS F, VARGA Z, KRUMREY M, NIEU-WLAND R. Innovation in detection of microparticles and exosomes. J Thromb Haemost 2013; 11 Suppl 1: 36-45.
- 60) VAN DER POL E, COUMANS FA, GROOTEMAAT AE, GARDINER C, SARGENT IL, HARRISON P, STURK A, VAN LEEUWEN TG, NIEUWLAND R. Particle size distribution of exosomes and microvesicles determined by transmission electron microscopy, flow cytometry, nanoparticle tracking analysis, and resistive pulse sensing. J Thromb Haemost 2014; 12: 1182-1192.
- 61) Tatischeff I, Larouet E, Falcon-Perez JM, Turpin PY, Kruglik SG. Fast characterisation of cell-derived extracellular vesicles by nanoparticles tracking analysis, cryo-electron microscopy, and Raman tweezers microspectroscopy. J Extracell Vesicles 2012;1.
- 62) CAMUSSI G, DEREGIBUS MC, BRUNO S, CANTALUPPI V, BIANCONE L. Exosomes/microvesicles as a mechanism of cell-to-cell communication. Kidney Int 2010; 78: 838-848.

- 63) VALADI H, EKSTROM K, BOSSIOS A, SJOSTRAND M, LEE JJ, LOTVALL JO. Exosome-mediated transfer of mR-NAs and microRNAs is a novel mechanism of genetic exchange between cells. Nat Cell Biol 2007; 9: 654-659.
- 64) SKOG J, WURDINGER T, VAN RIJN S, MEJJER DH, GAINCHE L, SENA-ESTEVES M, CURRY WT, JR., CARTER BS, KRICHEVSKY AM, BREAKEFIELD XO. Glioblastoma microvesicles transport RNA and proteins that promote tumour growth and provide diagnostic biomarkers. Nat Cell Biol 2008; 10: 1470-1476.
- 65) BARONI S, ROMERO-CORDOBA S, PLANTAMURA I, DUGO M, D'IPPOLITO E, CATALDO A, COSENTINO G, ANGELONI V, ROSSINI A, DAIDONE MG, IORIO MV. Exosome-mediated delivery of miR-9 induces cancer-associated fibroblast-like properties in human breast fibroblasts. Cell Death Dis 2016; 7: e2312.
- 66) BLIN J, FITZGERALD KA. Perspective: the RNA exosome, cytokine gene regulation and links to autoimmunity. Cytokine 2015; 74: 175-180.
- 67) EMANUELI C, SHEARN AI, ANGELINI GD, SAHOO S. Exosomes and exosomal miRNAs in cardiovascular protection and repair. Vascul Pharmacol 2015; 71: 24-30.
- 68) YUAN MJ, MAGHSOUDI T, WANG T. Exosomes mediate the intercellular communication after myocardial infarction. Int J Med Sci 2016; 13: 113-116.
- 69) KUWABARA Y, ONO K, HORIE T, NISHI H, NAGAO K, KINOSHITA M, WATANABE S, BABA O, KOJIMA Y, SHIZUTA S, IMAI M, TAMURA T, KITA T, KIMURA T. Increased microRNA-1 and microRNA-133a levels in serum of patients with cardiovascular disease indicate myocardial damage. Circ Cardiovasc Genet 2011; 4: 446-454.
- 70) EISSA S, MATBOLI M, ABOUSHAHBA R, BEKHET MM, SOLI-MAN Y. Urinary exosomal microRNA panel unravels novel biomarkers for diagnosis of type 2 diabetic kidney disease. J Diabetes Complications 2016; 30: 1585-1592.
- 71) EISSA S, MATBOLI M, BEKHET MM. Clinical verification of a novel urinary microRNA panal: 133b, -342 and -30 as biomarkers for diabetic nephropathy identified by bioinformatics analysis. Biomed Pharmacother 2016; 83: 92-99.
- CHISTIAKOV DA, SOBENIN IA, OREKHOV AN. Strategies to deliver microRNAs as potential therapeutics in the treatment of cardiovascular pathology. Drug Deliv 2012; 19: 392-405.
- LOYER X, VION AC, TEDGUI A, BOULANGER CM. Microvesicles as cell-cell messengers in cardiovascular diseases. Circ Res 2014; 114: 345-353.
- 74) HSU DH, PAZ P, VILLAFLOR G, RIVAS A, MEHTA-DAMANI A, ANGEVIN E, ZITVOGEL L, LE PECO JB. Exosomes as a tumor vaccine: enhancing potency through direct loading of antigenic peptides. J Immunother 2003; 26: 440-450.
- 75) LIU H, GAO W, YUAN J, WU C, YAO K, ZHANG L, MA L, ZHU J, ZOU Y, GE J. Exosomes derived from dendritic cells improve cardiac function via activation of CD4(+) T lymphocytes after myocardial infarction. J Mol Cell Cardiol 2016; 91: 123-133.

- 76) GARCIA NA, MONCAYO-ARLANDI J, SEPULVEDA P, DIEZ-JUAN A. Cardiomyocyte exosomes regulate glycolytic flux in endothelium by direct transfer of GLUT transporters and glycolytic enzymes. Cardiovasc Res 2016; 109: 397-408.
- 77) Yu B, Kim HW, Gong M, Wang J, MILLARD RW, Wang Y, Ashraf M, Xu M. Exosomes secreted from GA-TA-4 overexpressing mesenchymal stem cells serve as a reservoir of anti-apoptotic microRNAs for cardioprotection. Int J Cardiol 2015; 182: 349-360.
- 78) WESTERMEIER F, RIQUELME JA, PAVEZ M, GARRIDO V, DIAZ A, VERDEJO HE, CASTRO PF, GARCÍA L, LAVAN-DERO S. New molecular insights of insulin in diabetic cardiomyopathy. Front Physiol 2016; 7: 125.
- 79) GOUVEIA MC, VELLA JP, CAFEO FR, AFFONSO FONSECA FL, BACCI MR. Association between irisin and major chronic diseases: a review. Eur Rev Med Pharmacol Sci 2016; 20: 4072-4077.
- 80) ZAMPETAKI A, KIECHL S, DROZDOV I, WILLEIT P, MAYR U, PROKOPI M, MAYR A, WEGER S, OBERHOLLENZER F, BONO-RA E, SHAH A, WILLEIT J, MAYR M. Plasma microRNA profiling reveals loss of endothelial miR-126 and other microRNAs in type 2 diabetes. Circ Res 2010; 107: 810-817.
- 81) SANTOVITO D, DE NARDIS V, MARCANTONIO P, MANDOLINI C, PAGANELLI C, VITALE E, BUTTITTA F, BUCCI M, MEZZETTI A, CONSOLI A, CIPOLLONE F. Plasma exosome microRNA profiling unravels a new potential modulator of adiponectin pathway in diabetes: effect of glycemic control. J Clin Endocrinol Metab 2014; 99: E1681-1685.
- 82) Jansen F, Yang X, Baumann K, Przybilla D, Schmitz T, Flender A, Paul K, Alhusseiny A, Nickenig G, Werner N. Endothelial microparticles reduce ICAM-1 expression in a microRNA-222-dependent mechanism. J Cell Mol Med 2015; 19: 2202-2214.
- 83) WANG X, HUANG W, LIU G, CAI W, MILLARD RW, WANG Y, CHANG J, PENG T, FAN GC. Cardiomyocytes mediate anti-angiogenesis in type 2 diabetic rats through the exosomal transfer of miR-320 into endothelial cells. J Mol Cell Cardiol 2014; 74: 139-150.
- 84) CAPORALI A, MELONI M, NAILOR A, MITIC T, SHANTI-KUMAR S, RIU F, SALA-NEWBY GB, ROSE L, BESNIER M, KATARE R, VOELLENKLE C, VERKADE P, MARTELLI F, MADED-DU P, EMANUELI C. p75(NTR)-dependent activation of NF-kappaB regulates microRNA-503 transcription and pericyte-endothelial crosstalk in diabetes after limb ischaemia. Nat Commun 2015; 6: 8024.
- 85) HULSMANS M, HOLVOET P. MicroRNA-containing microvesicles regulating inflammation in association with atherosclerotic disease. Cardiovasc Res 2013; 100: 7-18.
- 86) Guay C, Menoud V, Rome S, Regazzi R. Horizontal transfer of exosomal microRNAs transduce apoptotic signals between pancreatic beta-cells. Cell Commun Signal 2015; 13: 17.
- 87) CHATURVEDI P, KALANI A, MEDINA I, FAMILTSEVA A, TYAGI SC. Cardiosome mediated regulation of MMP9 in

- diabetic heart: role of mir29b and mir455 in exercise. J Cell Mol Med 2015; 19: 2153-2161.
- 88) SHANTIKUMAR S, ANGELINI GD, EMANUELI C. Diabetes, microRNAs and exosomes: les liaisons dangereuses. J Mol Cell Cardiol 2014; 74: 196-198.
- 89) Nakano M, Nagaishi K, Konari N, Saito Y, Chikenji T, Mizue Y, Fujimiya M. Bone marrow-derived mesenchymal stem cells improve diabetes-induced cognitive impairment by exosome transfer into damaged neurons and astrocytes. Sci Rep 2016; 6: 24805.
- 90) Jansen F, Yang X, Proebsting S, Hoelscher M, Przybilla D, Baumann K, Schmitz T, Dolf A, Endl E, Franklin BS, Sinning JM, Vasa-Nicotera M, Nickenig G, Werner N. MicroRNA expression in circulating microvesicles predicts cardiovascular events in patients with coronary artery disease. J Am Heart Assoc 2014; 3: e001249.
- 91) Jansen F, Yang X, Hoelscher M, Cattelan A, Schmitz T, Proebsting S, Wenzel D, Vosen S, Franklin BS, Fleischmann BK, Nickenig G, Werner N. Endothelial microparticle-mediated transfer of MicroR-NA-126 promotes vascular endothelial cell repair via SPRED1 and is abrogated in glucose-damaged endothelial microparticles. Circulation 2013; 128: 2026-2038.
- 92) Geiger A, Walker A, Nissen E. Human fibrocyte-derived exosomes accelerate wound healing in genetically diabetic mice. Biochem Biophys Res Commun 2015; 467: 303-309.
- 93) Ni J, Lu H, Lu X, Jiang M, Peng Q, Ren C, Xiang J, Mei C, Li J. The evolving concept of physiological ischemia training vs. ischemia preconditioning. J Biomed Res 2015; 29: 445-450.
- 94) ZHANG Y, LIU YJ, LIU T, ZHANG H, YANG SJ. Plasma microRNA-21 is a potential diagnostic biomarker of acute myocardial infarction. Eur Rev Med Pharmacol Sci 2016; 20: 323-329.
- Gallo S, Sala V, Gatti S, Crepaldi T. Cellular and molecular mechanisms of HGF/Met in the cardiovascular system. Clin Sci (Lond) 2015; 129: 1173-1193.
- 96) CHAIKRIANGKRAI K, POLSANI V, WEI L, KLEIMAN N, CHANG SM. Stenting of a left main coronary artery compressed by a dilated main pulmonary artery. Catheter Cardiovasc Interv 2013; 82: E684-687.
- 97) LOPATINA T, GAI C, DEREGIBUS MC, KHOLIA S, CAMUSSI G. Cross talk between cancer and mesenchymal stem cells through extracellular vesicles carrying nucleic acids. Front Oncol 2016; 6: 125.
- 98) Prattichizzo F, Giuliani A, De Nigris V, Pujadas G, Ceka A, La Sala L, Genovese S, Testa R, Procopio AD, Olivieri F, Ceriello A. Extracellular microRNAs and endothelial hyperglycaemic memory: a therapeutic opportunity? Diabetes Obes Metab 2016; 18: 855-867.
- 99) O'Neill S, Bohl M, Gregersen S, Hermansen K, O'Driscoll L. Blood-based biomarkers for metabolic syndrome. Trends Endocrinol Metab 2016; 27: 363-374.

- 100) KISHORE R, GARIKIPATI VN, GUMPERT A. Tiny shuttles for information transfer: exosomes in cardiac health and disease. J Cardiovasc Transl Res 2016; 9: 169-175.
- 101) SAHOO S, LOSORDO DW. Exosomes and cardiac repair after myocardial infarction. Circ Res 2014; 114: 333-344.
- 102) GRAY WD, FRENCH KM, GHOSH-CHOUDHARY S, MAX-WELL JT, BROWN ME, PLATT MO, SEARLES CD, DAVIS ME. Identification of therapeutic covariant microRNA clusters in hypoxia-treated cardiac progenitor cell exosomes using systems biology. Circ Res 2015; 116: 255-263.
- 103) Kervadec A, Bellamy V, El Harane N, Arakelian L, Vanneaux V, Cacciapuoti I, Nemetalla H, Périer MC, Toeg HD, Richart A, Lemitre M, Yin M, Loyer X, Larghero J, Hagège A, Ruel M, Boulanger CM, Silvestre JS, Menasché P, Renault NK. Cardiovascular progenitor-derived extracellular vesicles recapitulate the beneficial effects of their parent cells in the treatment of chronic heart failure. J Heart Lung Transplant 2016; 35: 795-807.
- 104) Yu B, Gong M, Wang Y, MILLARD RW, PASHA Z, YANG Y, ASHRAF M, Xu M. Cardiomyocyte protection by GATA-4 gene engineered mesenchymal stem cells is partially mediated by translocation of miR-221 in microvesicles. PLoS One 2013; 8: e73304.
- 105) IBRAHIM AG, CHENG K, MARBAN E. Exosomes as critical agents of cardiac regeneration triggered by cell therapy. Stem Cell Reports 2014; 2: 606-619
- 106) LAI RC, ARSLAN F, LEE MM, SZE NS, CHOO A, CHEN TS, SALTO-TELLEZ M, TIMMERS L, LEE CN, EL OAKLEY RM, PASTERKAMP G, DE KLEIJN DP, LIM SK. Exosome secreted by MSC reduces myocardial ischemia/reperfusion injury. Stem Cell Res 2010; 4: 214-222.
- 107) Arslan F, Lai RC, Smeets MB, Akeroyd L, Choo A, Aguor EN, Timmers L, Van Rijen HV, Doevendans PA, Pasterkamp G, Lim SK, De Kleijn DP. Mesenchymal stem cell-derived exosomes increase ATP levels, decrease oxidative stress and activate PI3K/Akt pathway to enhance myocardial viability and prevent adverse remodeling after myocardial ischemia/reperfusion injury. Stem Cell Res 2013; 10: 301-312.
- 108) VICENCIO JM, YELLON DM, SIVARAMAN V, DAS D, BOI-DOKU C, ARJUN S, ZHENG Y, RIQUELME JA, KEAR-NEY J, SHARMA V, MULTHOFF G, HALL AR, DAVIDSON SM. Plasma exosomes protect the myocardium from ischemia-reperfusion injury. J Am Coll Cardiol 2015; 65: 1525-1536.
- 109) LIBBY P, RIDKER PM, MASERI A. Inflammation and atherosclerosis. Circulation 2002; 105: 1135-1143.
- 110) FLORE R, PONZIANI FR, TINELLI G, ARENA V, FONNESU C, NESCI A, SANTORO L, TONDI P, SANTOLIQUIDO A. New modalities of ultrasound-based intima-media thickness, arterial stiffness and non-coronary vascular calcifications detection to assess cardiovascular risk. Eur Rev Med Pharmacol Sci 2015; 19: 1430-1441.

- 111) ZHAN R, LENG X, LIU X, WANG X, GONG J, YAN L, WANG L, WANG Y, WANG X, QIAN LJ. Heat shock protein 70 is secreted from endothelial cells by a non-classical pathway involving exosomes. Biochem Biophys Res Commun 2009; 387: 229-233.
- 112) CHEN X, SUN Z, Du X, LIU C, LIU Y, WU L. Study on the relationship between heat shock protein 70 and toll-like receptor-4 of monocytes. J Huazhong Univ Sci Technolog Med Sci 2004; 24: 560-562.
- 113) TAIBI F, METZINGER-LE MEUTH V, MASSY ZA, METZINGER L. miR-223: an inflammatory oncomiR enters the cardiovascular field. Biochim Biophys Acta 2014; 1842: 1001-1009.
- 114) RAUTOU PE, LEROYER AS, RAMKHELAWON B, DEVUE C, DUFLAUT D, VION AC, NALBONE G, CASTIER Y, LESECHE G, LEHOUX S, TEDGUI A, BOULANGER CM. Microparticles from human atherosclerotic plaques promote endothelial ICAM-1-dependent monocyte adhesion and transendothelial migration. Circ Res 2011; 108: 335-343.
- 115) BOON RA, HORREVOETS AJ. Key transcriptional regulators of the vasoprotective effects of shear stress. Hamostaseologie 2009; 29: 39-40, 41-33.
- 116) HERGENREIDER E, HEYDT S, TREGUER K, BOETTGER T, HORREVOETS AJ, ZEIHER AM, SCHEFFER MP, FRANGAKIS AS, YIN X, MAYR M, BRAUN T, URBICH C, BOON RA, DIMMELER S. Atheroprotective communication between endothelial cells and smooth muscle cells through miRNAs. Nat Cell Biol 2012; 14: 249-256.
- 117) CHENG Y, LIU X, YANG J, LIN Y, XU DZ, LU Q, DE-ITCH EA, HUO Y, DELPHIN ES, ZHANG C. MicroR-NA-145, a novel smooth muscle cell phenotypic marker and modulator, controls vascular neointimal lesion formation. Circ Res 2009; 105: 158-166.
- 118) HALKEIN J, TABRUYN SP, RICKE-HOCH M, HAGHIKIA A, NGUYEN NQ, SCHERR M, CASTERMANS K, MALVAUX L, LAMBERT V, THIRY M, SLIWA K, NOEL A, MARTIAL JA, HILFIKER-KLEINER D, STRUMAN I. MICRORNA-146a is a therapeutic target and biomarker for peripartum cardiomyopathy. J Clin Invest 2013; 123: 2143-2154.
- 119) YANG Y, RODRIGUEZ JE, KITSIS RN. A microRNA links prolactin to peripartum cardiomyopathy. J Clin Invest 2013; 123: 1925-1927.
- 120) HALKEIN J, DE WINDT LJ. miR-223: sailing to terra incognita for microRNAs in platelets. Thromb Haemost 2013; 110: 1112-1113.
- 121) KANEIDER NC, KASER A, TILG H, RICEVUTI G, WIEDER-MANN CJ. CD40 ligand-dependent maturation of human monocyte-derived dendritic cells by activated platelets. Int J Immunopathol Pharmacol 2003; 16: 225-231.
- 122) SADALLAH S, EKEN C, MARTIN PJ, SCHIFFERLI JA. Microparticles (ectosomes) shed by stored human platelets downregulate macrophages and modify the development of dendritic cells. J Immunol 2011; 186: 6543-6552.

- 123) Weber A, Koppen HO, Schror K. Platelet-derived microparticles stimulate coronary artery smooth muscle cell mitogenesis by a PDGF-independent mechanism. Thromb Res 2000; 98: 461-466.
- 124) LIU ML, SCALIA R, MEHTA JL, WILLIAMS KJ. Cholesterol-induced membrane microvesicles as novel carriers of damage-associated molecular patterns: mechanisms of formation, action, and detoxification. Arterioscler Thromb Vasc Biol 2012; 32: 2113-2121.
- 125) Hoyer FF, Giesen MK, Nunes Franca C, Lutjohann D, Nickenig G, Werner N. Monocytic microparticles promote atherogenesis by modulating inflammatory cells in mice. J Cell Mol Med 2012; 16: 2777-2788.
- 126) Mastronardi ML, Mostefai HA, Soleti R, Agouni A, Martinez MC, Andriantsitohaina R. Microparticles from apoptotic monocytes enhance nitrosative stress in human endothelial cells. Fundam Clin Pharmacol 2011; 25: 653-660.